

## Dr. rer. nat. Sven Timo Stripp

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Wittelsbacherstrasse 16

10707 Berlin



### PERSONAL INFORMATION

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Born 1980 in Wesel, Germany

Married, two children (\*2009 and \*2012)

### RESEARCH EXPERIENCE

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**2010 – today**

**Freie Universität Berlin**

Senior scientist/ group leader (h-index = 17, > 1.100 citations)

Spectroscopic developments for the analysis of gas-processing metalloenzymes.

Department of Physics, Experimental Molecular Biophysics (Mentor: Prof. Dr. J. Heberle)

### EDUCATION

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**2007 – 2010**

**Ruhr-Universität Bochum**

*Dissertation in Biology, degree: Dr. rer. nat. (summa cum laude)*

“Molecular Background of Oxygen Sensitivity in [FeFe] hydrogenases.”

Faculty of Biology and Biotechnology, Photobiotechnology.

Prof. Dr. T. Happe and Prof. Dr. E. Hofmann

**2001 – 2007**

**Ruhr-Universität Bochum**

*Studies of Chemistry and Biology, degree: diploma (1.7)*

“Optimierung der Expression eines Proteins aus *C. reinhardtii* in *C. acetobutylicum*.”

Faculty of Biology and Biotechnology, Photobiotechnology.

Prof. Dr. T. Happe and Prof. Dr. K. Störtkuhl

## RESEARCH PROFILE (*brief*)

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### Main Research Interests

- metalloenzymes and biocatalysis
- enzymatic activation of small molecules, in particular H<sub>2</sub>, N<sub>2</sub>, O<sub>2</sub>, CO<sub>2</sub>/ CO (context: green chemistry/ fuel cells, energy conservation, greenhouse gases)
- proton-coupled electron transfer (PCET) and proton transfer in biology
- interplay of protein fold and cofactor

### Spectroscopic Expertise

- attenuated total reflection Fourier-transform IR spectroscopy (ATR FTIR) with focus on novel techniques for *in situ* modification of sample matter
- protein film electrochemistry and spectro-electrochemistry
- surface-enhanced IR absorption spectroscopy (SEIRAS)
- time-resolved quantum cascade laser (QCL) IR spectroscopy
- molecular biology

### Collaboration Partners (active)

- Dr. Ulf-Peter Apfel (Ruhr-Universität Bochum/ Fraunhofer UMSICHT)
- Prof. Dr. R. David Britt (University of California Davis)
- Prof. Dr. Thomas Happe (Ruhr-Universität Bochum)
- Dr. Michael Haumann (Freie Universität Berlin)
- Prof. Dr. Joachim Heberle (Freie Universität Berlin)
- Dr. Seigo Shima (MPI for Terrestrial Microbiology, Marburg)
- Dr. Basem Soboh (Freie Universität Berlin)
- Prof. Dr. Kylie Vincent (Oxford University)
- Prof. Dr. Jiangyun Wang (Chinese Academy of Sciences, Beijing)
- Dr. Ingo Zebger (Technische Universität Berlin)

### Research Stays

- Hamburg (Deutsches Elektronen-Synchrotron, Dr. M. Haumann),
- Uppsala, SWE (Ångström Laboratory)
- Oxford, UK (Inorganic Chemistry Laboratory, Prof. Dr. F. Armstrong)

### Other

- Member of the “Deutsche Gesellschaft für Biophysik” (DGfB)
- Referee for international renowned journals such as “Proceedings of the National Academy of Sciences U.S.A.” (PNAS), “Journal of the American Chemical Society” (JACS), etc.
- Representative of mid-level faculty e.g. in search committees, dissertation defenses, etc.
- Organizer of the “Hydrogenase Minisymposium” in 2018 (Berlin)
- Administrator at [twitter.com/H2ASE\\_NEWS](https://twitter.com/H2ASE_NEWS)

## CONFERENCES AND SYMPOSIA (*selected*)

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**12<sup>th</sup> Int. Conference on Hydrogenases** (2019, Lisbon, Portugal):

“Proton Transfer Dynamics in [FeFe]-hydrogenases” (*invited talk*)

**Biennial Meeting of the German Biophysical Society DGfB** (2018, Düsseldorf)

“Proton Transfer across the Protein Fold is Coupled to Redox Changes at the Catalytic Metal Center of Hydrogen-producing Enzymes” (contributed talk)

**1<sup>st</sup> Hydrogenase Minisymposium** (2018, Berlin) (*organizer*)

**43<sup>rd</sup> Int. Conference on Coordination Chemistry ICC** (2018, Sendai, Japan):

“The Molecular Proceedings of Biological Hydrogen Turnover” (*invited talk*)

**2<sup>nd</sup> Progress Meeting of DFG Priority Program 1927** (2018, Potsdam):

„Biospectroscopy on gas-processing Enzymes” (*invited talk*)

**Gordon Research Conference “Metals in Biology”** (2018, Ventura, USA):

“Proton-coupled Electron Transfer at the Catalytic Cofactor of Hydrogenases”

**17<sup>th</sup> European Conference on the Spectroscopy of Biological Molecules ECSBM** (2017, Amsterdam, NL):

“Proton-coupled Electron Transfer at the Catalytic Cofactor of Hydrogenases” (contributed talk)

**Gordon Research Conference “Cell Biology of Metals”** (2017, Mt. Snow, USA):

“Proton-coupled Electron Transfer at the Catalytic Cofactor of Hydrogenases”

**11<sup>th</sup> Int. Conference on Hydrogenases** (2016, Marseilles, FR):

“Stepwise Isotope Editing of [FeFe] Hydrogenases” (contributed talk)

**1<sup>st</sup> International Solar Fuels conference ISF** (2015, Uppsala, SWE):

“The Yin and Yang of Hydrogen Turnover” (contributed talk)

## EXTERNAL FUNDING

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**Focus area Functional Materials at the Nanoscale** ([nanoscale.fu-berlin.de](http://nanoscale.fu-berlin.de))

Project support in 2015 (€ 22.000) and 2013 (€ 30.000)

**Deutscher Akademischer Austauschdienst (DAAD)**

In 2018, I received a travel grant for an invited talk in Japan.

**Deutsche Forschungsgemeinschaft (DFG)**

I applied for funding of my position via the Heisenberg Program (grant proposals STR 1554 1-2 and 1-2)

Furthermore, I assisted in the fabrication and correction of several grant applications, most recently for the Cluster of Excellence UniSysCat (project E2).

## TEACHING

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### SS 2012 – 2018

- “Advanced Biophysics” (lecture for Master with Prof. J. Heberle)

The lecture is designed to introduce spectroscopic/ physical techniques of particular relevance in molecular life science to an audience of physics students with a background in biophysics or biochemistry (see “Biophysik für Bachelor” below). Topics include absorption-, fluorescence-, and scattering spectroscopy in the UV, visible and IR regime; spectroscopy with linear and circular polarized light; surface-enhanced Raman and IR spectroscopy; time-resolved approaches; electron paramagnetic resonance (EPR) and nuclear magnetic resonance (NMR); diffraction with X-rays, Neutrons, and electrons; crystallization and protein crystallography; theoretical methods: MD simulations, Poisson-Boltzmann, QM/MM.

- “Advanced Laboratory Course in Biophysics” (lab course for Master and admin)

The lab course introduces a couple of spectroscopic/ physical techniques in a hands-on approach. The focus is on membrane proteins. At the example of bacteriorhodopsin, steady state (FTIR, RR) and time-resolved spectroscopy (UV/ vis and QCL IR flash-photolysis, step-scan FTIR) is performed. Furthermore, amino acid residues and simple peptides in KBr pellets are probed by FTIR spectroscopy. The band assignment proceeds via both tabulated values and DFT calculations. Purple membrane patches are investigated by AFM and SMFS. Eventually, the elastic properties and phase transitions of a lipid layer are probed in a Langmuir-Blodgett trough.

### WS/SS 2015 – 2018

- “Physik für die Grundschule” (lecture for Bachelor with J. Fahndrich and seminar)

Teacher training course for students of natural sciences. Making use of educational examples related to everyday life, methods and basic principles of physics are introduced. Topics include electricity and magnetism; optics and (visible) light; classical mechanics; liquids, gases, and temperature; acoustics. In the seminar, simple experiments are employed to demonstrate these principles and deepen the understanding.

### WS 12/13 – 17/18

- “Biophysik für Bachelor” (lecture for Bachelor with Prof. J. Heberle and seminar)

Proteins and in particular membrane proteins are introduced to students of physics with the aim to invoke an understanding of structure, dynamics, and function of biological macromolecules. Topics include structure and folding of proteins; hydrophobic effect and protein electrostatics; temperature and protein dynamics; basics of MD calculations; Mitchells’ theory of energy conservation (e.g. rhodopsins, photosynthetic and mitochondrial transport chain, ATP synthase); light- and voltage-gated channels; induction and propagation of action potentials; magneto reception.

**WS/SS 2016 – 2018**

- “Biophysik für Biochemiker” (lecture for Bachelor with R. Schlesinger and lab course)

The lectures introduces basic concepts of vibrational and UV/ vis spectroscopy to students of biochemistry. Topics include IR and Raman spectroscopy; mathematics of Fourier-transformation; scattering and resonance; time-resolved approaches (rapid-scan, step-scan, pump-probe). For the lab course, students of biochemistry perform a selected set of experiments on bacteriorhodopsin, including FTIR and Resonance Raman spectroscopy, step-scan FTIR, and UV/ vis flash photolysis.

**WS/SS 2012 – 2014**

- “Physikalisches Grundpraktikum und Grundlagen der Mess- und Labortechnik” (lab course for Ba.)

Introduction to the fundamental techniques of quantitative experimental and scientific methods in physics, i.e. measurement methods and techniques; documentation; mathematical-statistical und practical evaluation (error calculations); critical discussion and scientific conclusion; written report and presentation. I did course for students of physics, chemistry, and medicine.

**Supervision of Students**

Bachelor	Master/ Diploma	PhD
Viktor Eichmann (2017)	Konstantin Laun (2018)	Moritz Senger (2014 - 2017)
Konstantin Laun (2016)	Moritz Senger (2012)	
Iuliia Baranova (2016)		
Olga Shulenina (2015)		

**Research Seminars and Public Outreach**

WS/SS 2012 – 2018	“Function and Dynamics of Photoreceptors” (Research seminar)
WS/SS 2014 – 2018	“Metalloenzymes” (Research seminar)
SS 2012 – 2018	“Lange Nacht der Wissenschaft“ and “inFU:tage“

## PUBLICATION LIST

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1. Optimized over-expression of [FeFe] hydrogenases with high specific activity in *Clostridium acetobutylicum*. von Abendroth G, **Stripp ST**, Silakov A, Croux C, Soucaille P, Girbal L, Happe T\*. *Int. J. Hydrogen Energy*. 2008; 33(21): 6076-81.
2. The structure of the active site H-cluster of [FeFe] hydrogenase from the green alga *Chlamydomonas reinhardtii* studied by X-ray absorption spectroscopy. **Stripp ST**, Sanganas O, Happe T, Haumann M\*. *Biochemistry*. 2009; 48(22): 5042-9.
3. Immobilization of the [FeFe]-hydrogenase CrHydA1 on a gold electrode: design of a catalytic surface for the production of molecular hydrogen. Krassen H, **Stripp ST**, von Abendroth G, Ataka K, Happe T, Heberle J\*. *J. Biotechnol.* 2009; 142(1): 3-9.
4. How oxygen attacks [FeFe] hydrogenases from photosynthetic organisms. **Stripp ST**, Goldet G, Brandmayr C, Sanganas O, Vincent KA, Haumann M, Armstrong FA, Happe T\*. *Proc. Natl. Acad. Sci. U S A*. 2009; 106(41): 17331-6.
5. Electrochemical kinetic investigations of the reactions of [FeFe]-hydrogenases with CO and O<sub>2</sub>: comparing the importance of gas tunnels and active-site electronic/redox effects. Goldet G, Brandmayr C, **Stripp ST**, Happe T, Cavazza C, Fontecilla-Camps JC, Armstrong FA\*. *J. Am. Chem. Soc.* 2009; 131(41): 14979-89.
6. How algae produce hydrogen - news from the photosynthetic hydrogenase. **Stripp ST**, Happe T\*. *Dalton Trans.* 2009; 45: 9960-9.
7. Multiple ferredoxin isoforms in *Chlamydomonas reinhardtii* – Their role under stress conditions and biotechnological implications. Winkler M, Hemschemeier A, Jacobs J, **Stripp ST**, Happe T\*. *Eur. J. Cell Biol.* 2010; 89(12): 998-1004.
8. Molecular Background of Oxygen Sensitivity in [FeFe]-hydrogenases. **Sven ST\***. 2011. Saarbrücken: Südwestdeutscher Verlag für Hochschulschriften.
9. The [FeFe]-hydrogenase maturation protein HydF contains a H-cluster like [4Fe4S]-2Fe site. Czech I, **Stripp ST**, Sanganas O, Leidel N, Happe T, Haumann M\*. *FEBS Lett.* 2011; 585(1): 225-30.
10. Tailor-Made Modification of a Gold Surface for the Chemical Binding of a High-Activity [FeFe] Hydrogenase. Krassen H, **Stripp ST**, Böhm N, Berkessel A, Happe T, Ataka K, Heberle J\*. *Eur. J. Inorg. Chem.* 2011: 1138–1146.

11. Formaldehyde - a rapid and reversible inhibitor of hydrogen production by [FeFe]-hydrogenases. Wait AF, Brandmayr C, **Stripp ST**, Cavazza C, Fontecilla-Camps JC, Happe T, Armstrong FA\*. J. Am. Chem. Soc. 2011; 133(5): 1282-5.
12. [NiFe]-hydrogenase maturation: isolation of a HypC-HypD complex carrying diatomic CO and CN<sup>-</sup> ligands. Soboh B, **Stripp ST**, Muhr E, Granich C, Braussemann M, Herzberg M, Heberle J, Gary Sawers R\*. FEBS Lett. 2012; 586(21): 3882-7.
13. Surface-enhanced infrared absorption spectroscopy (SEIRAS) to probe monolayers of membrane proteins. Ataka K, **Stripp ST**, Heberle J\*. Biochim. Biophys. Acta. 2013;1828(10):2283-93.
14. The [NiFe]-hydrogenase accessory chaperones HypC and HybG of Escherichia coli are iron- and carbon dioxide-binding proteins. Soboh B, **Stripp ST**, Bielak C, Lindenstrauß U, Braussemann M, Javid M, Hallensleben M, Granich C, Herzberg M, Heberle J, Sawers RG\*. FEBS Lett. 2013;587(16):2512-6.
15. HypD is the scaffold protein for Fe-(CN)<sub>2</sub>CO cofactor assembly in [NiFe]-hydrogenase maturation. **Stripp ST**, Soboh B, Lindenstrauß U, Braussemann M, Herzberg M, Nies DH, Sawers RG, Heberle J\*. Biochemistry. 2014; 52(19): 3289-96.
16. [NiFe]-hydrogenase maturation in vitro: analysis of the roles of the HybG and HypD accessory proteins. Soboh B\*, Lindenstrauß U, Granich C, Javed M, Herzberg M, Thomas C, **Stripp ST**. Biochem. J. 2014; 464(2): 169-77.
17. The influence of oxygen on [NiFe]-hydrogenase cofactor biosynthesis and how ligation of carbon monoxide precedes cyanation. **Stripp ST\***, Lindenstrauß U, Granich C, Sawers RG, Soboh B. PLoS One. 2014; 9(9): e107488. [\[open access\]](#)
18. Photosynthesis and hydrogen metabolism revisited. On the potential of light-driven hydrogen production in vitro. **Stripp ST**, Heberle J\*. Biohydrogen 2015 (pp. 211-238). Berlin, Boston: DE GRUYTER.
19. Identification of an Isothiocyanate on the HypEF Complex Suggests a Route for Efficient Cyanyl-Group Channeling during [NiFe]-Hydrogenase Cofactor Generation. **Stripp ST\***, Lindenstrauß U, Sawers RG, Soboh B. PLoS One. 2015; 10(7): e0133118. [\[open access\]](#)
20. Stepwise Isotope Editing of [FeFe]-Hydrogenases Exposes Cofactor Dynamics. Senger M, Mebs S, Duan J, Wittkamp F, Apfel UP, Heberle J, Haumann M, **Stripp ST\***. Proc. Natl. Acad. Sci. U S A. 2016; 113(30): 8454–59.
21. Wasserstoffkatalyse in Mikroalgen. Senger M and **Stripp ST\***. 2017 Nachrichten aus der Chemie. 2017; 65: 123-7. [\[open access\]](#) *(invited)*

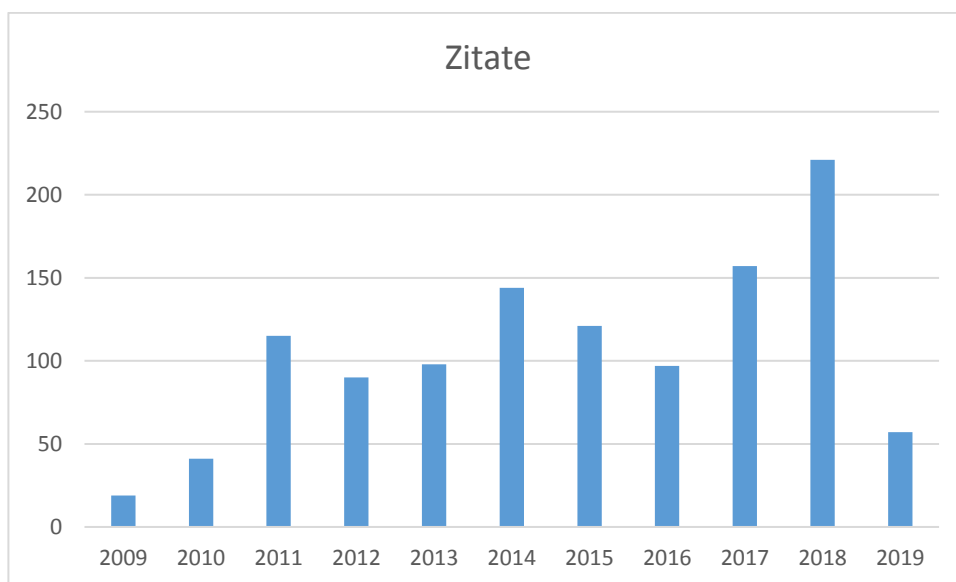
22. Proteolytic cleavage orchestrates cofactor insertion and protein assembly in [NiFe]-hydrogenase biosynthesis. Senger M, **Stripp ST**, Soboh B\*, J. Biol. Chem. 2017; 292(28): 11670-81. [\[open access\]](#)
23. Accumulating the Hydride State in the Catalytic Cycle of [FeFe]-Hydrogenases. Winkler M, Senger M, Duan J, Esselborn J, Wittkamp F, Hofmann E, Apfel UP, **Stripp ST\***, Happe T\* Nat. Comm. 2017; 8: 16115. [\[open access\]](#)
24. Bridging Hydride at Reduced H-Cluster Species in [FeFe]-Hydrogenases Revealed by Infrared Spectroscopy, Isotope Editing, and Quantum Chemistry. Mebs S\*, Senger M, Duan J, Wittkamp F, Apfel UP, Happe T, Winkler M, **Stripp ST\***, Haumann M\*. J. Am. Chem. Soc. 2017; 139: 12157–60.
25. Proton-Coupled Reduction of the Catalytic [4Fe-4S] Cluster in [FeFe]-Hydrogenases. Senger M, Laun K, Wittkamp F, Duan J, Happe T, Winkler M, Apfel UP\*, **Stripp ST\***. Angew. Chemie Int. Ed. 2017; 56 (52): 16503–06.
26. Protonengekoppelte Reduktion des katalytischen [4Fe-4S]-Zentrums in [FeFe]-Hydrogenasen. Senger M, Laun K, Wittkamp F, Duan J, Happe T, Winkler M, Apfel UP\*, **Stripp ST\***. Angew. Chem. 2017; 129 (52): 16728–32
27. Protonation and Reduction Dynamics at the Hydrogen-forming Cofactor of [FeFe]-Hydrogenases. Senger M, Mebs S, Duan J, Shulenina O, Laun K, Kertess L, Wittkamp F, Apfel UP, Happe T, Winkler M\*, Haumann M\*, **Stripp ST\***. Phys. Chem. Chem. Phys. 2018; 20: 3128–40. [\[open access\]](#) *(invited)*
28. Hydrogen and oxygen trapping at the H-cluster of [FeFe]-hydrogenase revealed by site-selective spectroscopy and QM/MM calculations. Mebs S, Kositzki R, Duan J, Senger M, Wittkamp F, Apfel UP, Happe T, **Stripp ST**, Winkler M\*, Haumann M\*. BBA - Bioenergetics 2018; 1859: 28–41. *(invited)*
29. Wasserstoffproduktion nach dem Vorbild der Nature. Apfel UP, **Stripp ST\***. GIT Laborfachzeitschrift 2018; 6: 28-29 [\[open access\]](#) *(invited)*
30. [FeFe]-hydrogenases: recent developments and future perspectives. Wittkamp F, Senger M, **Stripp ST**, Apfel UP\*. Chem. Comm. 2018; 54: 5934-5942 *(invited)*
31. Spectroscopical Investigations on the Redoxchemistry of [FeFe]-Hydrogenases in the Presence of Carbon Monoxide. Laun K, Mebs S, Duan J, Wittkamp F, Apfel UP, Happe T, Winkler M, Haumann M\*, **Stripp ST\***. MOLECULES 2018; 23: 1669 [\[open access\]](#) *(invited)*
32. The molecular proceedings of biological hydrogen turnover. Haumann M, **Stripp ST\***. Acc. Chem. Res. 2018; 51 (8): 1755 – 63 *(invited)*



33. Crystallographic and spectroscopic assignment of the proton transfer pathway in [FeFe]-hydrogenases. Duan J, Senger M, Esselborn J, Engelbrecht V, Wittkamp F, Apfel Ulf-Peter, Hofmann E, **Stripp ST**, Happe T\*, Winkler M\*. Nat. comm. 2018; 9: 4726 [[open access](#)]
34. Infrared Characterization of the Bidirectional O<sub>2</sub>-sensitive [NiFe]-hydrogenase from Escherichia coli. Senger M, Laun K, Soboh B, **Stripp ST\***. CATALYSTS 2018; 8: 530 [[open access](#)] (*invited*)
35. Differential Protonation at the Catalytic Six-Iron Cofactor of [FeFe]-Hydrogenases Revealed by <sup>57</sup>Fe Nuclear Resonance X-ray Scattering and Quantum Mechanics/Molecular Mechanics Analyses. Mebs S, Duan J, Wittkamp F, **Stripp ST**, Happe T, Apfel UP1, Winkler M, Haumann M\*. Inorg. Chem. 2019; 58 (5): 4000 – 16.
36. The Geometry of the Catalytic Active Site in [FeFe]-Hydrogenases is Determined by Hydrogen Bonding and Proton Transfer. Duan J, Mebs S, Senger M, Laun K, Wittkamp F, Heberle J, Happe T, Hofmann E, Apfel UP, Winkler M, Haumann M, **Stripp ST\***. ChemRxiv 2019. <https://doi.org/10.26434/chemrxiv.7756214.v1>

\* corresponding author

## PUBLICATION STATISTICS



Source: Google Scholar (issued 25.3.2019)

<https://scholar.google.com/citations?user=ZtfyzRoAAAAJ>

Zitate: 1174; h-index: 17; i10-index: 24

CERTIFICATES (*diploma*)

## RUHR-UNIVERSITÄT BOCHUM

FAKULTÄT FÜR BIOLOGIE

## ZEUGNIS

über die Diplomprüfung in Biologie

Herr Sven Timo **Stripp**

geb. am 07.01.1980 in wesel  
 hat am 14.09.2006 die Diplomprüfung in Biologie  
 vor dem Prüfungsausschuss für Biologie abgelegt.

Die Diplomarbeit

"Optimierung der heterologen Expression eines Proteins aus *Chlamydomonas reinhardtii*  
 in *Clostridium acetobutylicum*"

hat das Urteil gut (1,7)  
 erhalten.

Gutachter: Prof. Dr. T. Happe  
 Prof. Dr. K. Störtkuhl

Die Einzelergebnisse der mündlichen Prüfung sind:

Biologische Fachprüfung I:	Biochemie	sehr gut	(1,3)
Prüfer:	Prof. Dr. T. Happe		
Biologische Fachprüfung II:	Biotechnologie	sehr gut	(1,3)
Prüfer:	Prof. Dr. T. Happe		
Biologische Fachprüfung III:	Biophysik	gut	(2,0)
Prüfer:	PD Dr. J. Schlitter		
Außerbiol. Wahlpflichtfach:	Neuroinformatik	sehr gut	(1,0)
Prüfer:	Prof. Dr. G. Schöner		

Herr Sven Timo Stripp  
 hat die Diplomprüfung bestanden mit der Gesamtnote: sehr gut (1,5)

Bochum, den 08.11.2006

Der Vorsitzende des Prüfungsausschusses



(Prof. Dr. F. Narberhaus)

**CERTIFICATES (dissertation)**

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**RUHR-UNIVERSITÄT BOCHUM**  
**FAKULTÄT FÜR BIOLOGIE**  
**UND BIOTECHNOLOGIE**  
INTERNATIONALE GRADUIERTENSCHULE  
BIOWISSENSCHAFTEN

**Sven Timo Stripp,**

geboren am 07.01.1980 in Wesel, Deutschland

hat in einem ordnungsgemäßen Promotionsverfahren der Internationalen Graduiertenschule Biowissenschaften den Grad eines Doktors der Naturwissenschaften erlangt.

Titel der Dissertation:

**“Molekulare Grundlagen der Sauerstoffsensitivität  
von [FeFe]-Hydrogenasen”**

Promotionskommission:

Vorsitzender:	Prof. Dr. K. Gerwert
1. Referent:	Prof. Dr. T. Happe
2. Referent:	Prof. Dr. E. Hofmann
3. Prüferin:	Prof. Dr. N. Frankenberg-Dinkel


Ergebnis der Dissertation: **sehr gut (magna cum laude)**

Ergebnis der Disputation : **sehr gut (magna cum laude)**

Gesamtergebnis: **mit Auszeichnung (summa cum laude)**

Bochum, den 05.05.2010



  
Prof. Dr. F. Narberhaus  
(Dekan der Fakultät für Biologie  
und Biotechnologie)

**CERTIFICATES (teaching)**

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**Berlin, 19.07.2018**

**Bestätigung durchgeführter Lehrveranstaltungen**

Sehr geehrter Herr Stripp,

hiermit bestätigen wir Ihnen, dass Sie folgende Lehrveranstaltungen selbständig durchgeführt haben:

Semester	Stunden/ Woche	Titel der Veranstaltung	Art der Lehrdurchführung
SS 11	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
SS 12	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
	3,5	Physikalisches Grundpraktikum für Bachelor	Praktikumsbetreuung
WS 12/13	2	Physikalisches Grundpraktikum für Bachelor	Praktikumsbetreuung
SS 13	2	Physikalisches Grundpraktikum für Bachelor	Praktikumsbetreuung
	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
WS 13/14	4	Biophysik für Bachelor	Übungsgruppenbetreuung
SS 14	2	Physikalisches Grundpraktikum für Bachelor	Praktikumsbetreuung
	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
WS 14/15	2	Physik für die Grundschule 1	Übungsgruppenbetreuung
SS 15	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
	2	Physik für die Grundschule 2	Übungsgruppenbetreuung
WS 15/16	2	Biophysik für Bachelor	Übungsgruppenbetreuung
	1	Photobiophysics and Photosynthesis	Vorlesung
	1	Photobiophysics and Photosynthesis	Übungsgruppenbetreuung
SS 16	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
WS 16/17	2	Biophysik für Bachelor	Übungsgruppenbetreuung
	1	Production and biophysical analysis of selected membrane proteins Part I	Vorlesung
	2	Physik für die Grundschule 1	Übungsgruppenbetreuung

SS 17	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
	1	Production and Biophysical Analysis of Selected Membrane Proteins Part II	Vorlesung
	2	Physik für die Grundschule 2	Übungsgruppenbetreuung
WS 17/18	2	Biophysik für Bachelor	Übungsgruppenbetreuung
	2	Physik für die Grundschule 1	Übungsgruppenbetreuung
	1	Production and biophysical analysis of selected membrane proteins Part I	Vorlesung
SS 18	2	Advanced Laboratory Course in Biophysics for Master Students	Praktikumsbetreuung
	2	Physik für die Grundschule 2	Übungsgruppenbetreuung

Neben der dokumentierten Lehrtätigkeit in den Seminaren, Übungen und Praktika im Bereich Experimentelle Molekulare Biophysik war Herr Dr. Sven Timo Stripp von WS/SS 2011 bis WS/SS 2017 durchgehend an den Vorlesungen der Biophysik für Bachelor- und Master-Studenten beteiligt. Das beinhaltet konzeptionelle Beiträge zum Inhalt der Vorlesung und Klausuren, sein Einsatz als Prüfungsbeisitzer und als Dozent mit 50%-iger Übernahme der Vorlesung "Photobiophysics & Photosynthesis" im WS 15/16.

Mit freundlichen Grüßen



Prof. Dr. Paul Fumagalli  
(Studiendekan)